



Studies in marine macrolide synthesis: stereocontrolled synthesis of a C21–C34 subunit of the aplyronines

Ian Paterson,* Simon B. Blakey and Cameron J. Cowden

University Chemical Laboratory, Lensfield Road, Cambridge CB2 1EW, UK

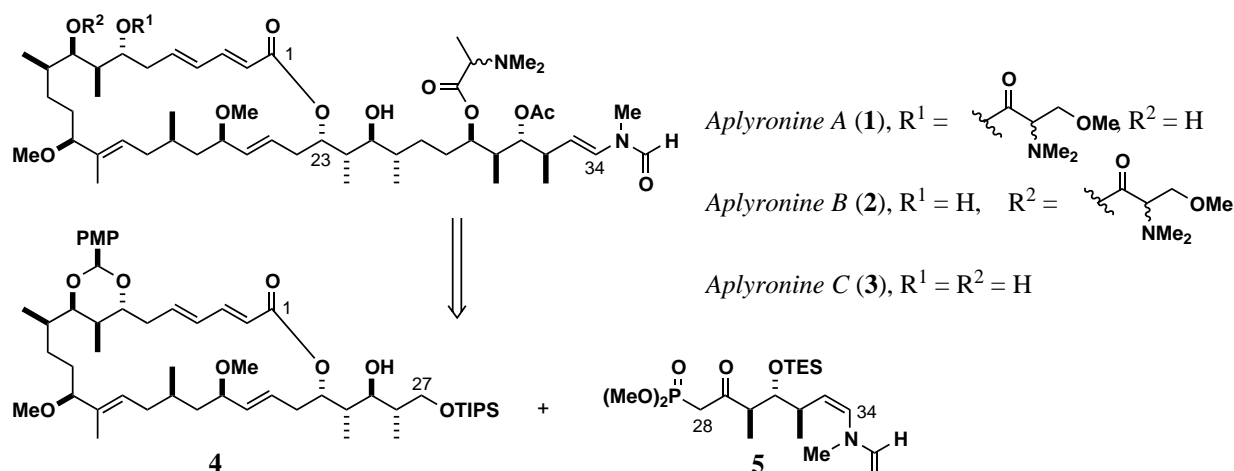
Received 27 May 2002; accepted 24 June 2002

Abstract—The C21–C34 subunit **27** of the aplyronines, containing eight stereocentres and a terminal *N*-methyl-*N*-vinylformamide moiety, was prepared using the Horner–Wadsworth–Emmons coupling of β -ketophosphonate **5** with aldehyde **19**. The two stereotetrad sequences were constructed by chiral ketone aldol reactions, while the *N*-methyl-*N*-vinylformamide was introduced using a novel Wittig olefination. © 2002 Elsevier Science Ltd. All rights reserved.

Aplyronine A (**1**, Scheme 1) is an unusual 24-membered marine macrolide which was first isolated in 1993, along with two congeners, aplyronines B (**2**) and C (**3**), from the Japanese sea hare *Aplysia kurodai* by Yamada and co-workers.¹ The aplyronines displayed potent cytotoxicity in vitro against a range of cancer cell lines including P388 leukaemia, Lewis lung carcinoma and B16 melanoma. Furthermore, aplyronine A exhibited pronounced in vivo activity against a range of tumour cells in xenograft experiments.^{1,2} Aplyronine A has been shown to function as a novel actin depolymerising agent,³ by accelerating fibrous actin depolymerisation and sequestering globular actin. Recently, an actin-dependent cell cycle checkpoint that ensures the proper

orientation of microtubule spindles during metaphase has been uncovered by Gachet and Hyams and co-workers,⁴ and although the biochemical pathway responsible for this checkpoint is still undefined, this may be linked to the antimitotic activity of the aplyronines.

The potent antitumour activity and novel actin-binding properties of the aplyronines has led to interest in evaluating their chemotherapeutic potential,³ as well as attracting synthetic attention towards this unique group of marine macrolides.^{2,5–7} Notably, the Yamada group established the absolute configuration by completing the first total synthesis of aplyronines A–C.² We have



Scheme 1.

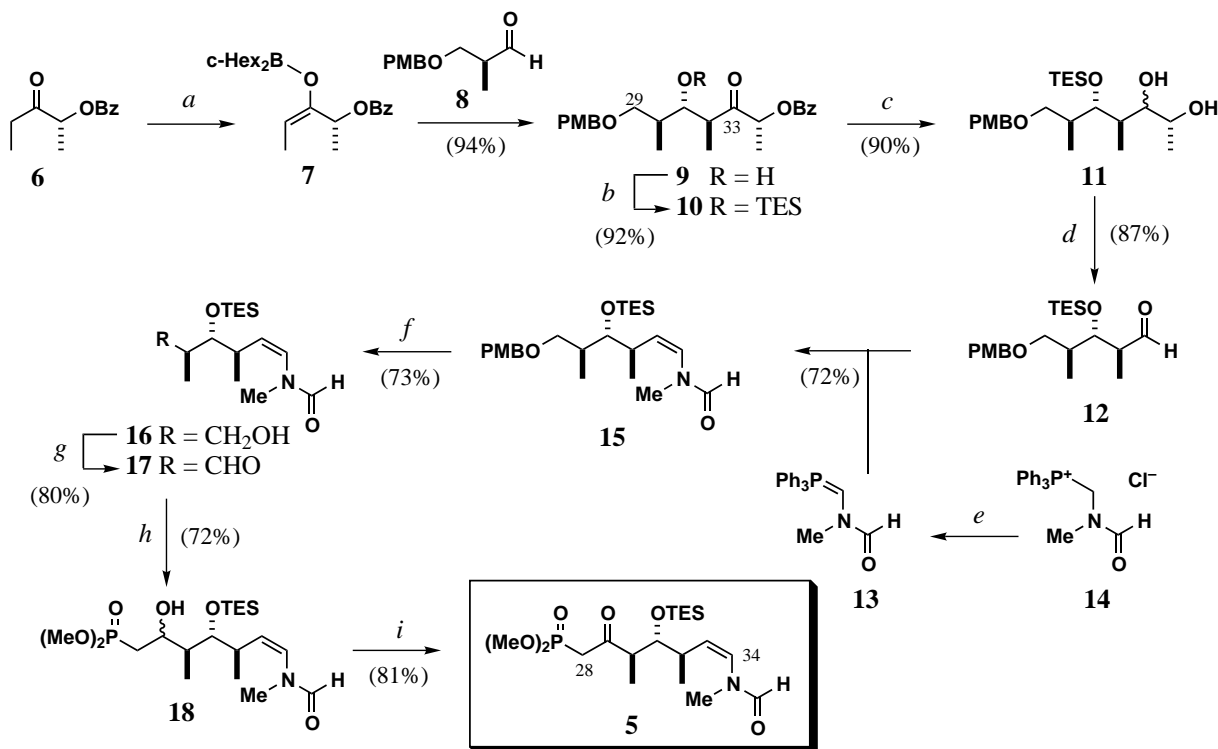
* Corresponding author. Fax: +44-1223-336362; e-mail: ip100@cus.cam.ac.uk

adopted an alternative strategy for the stereocontrolled synthesis of the aplyronines, which is potentially shorter and relies upon a key Horner–Wadsworth–Emmons (HWE) coupling of a suitable C27 aldehyde, derived from the previously described macrolide **4**,⁵ with the β -ketophosphonate **5** for elaboration of the side chain. We now report a synthesis of the C28–C34 phosphonate **5** and demonstrate its use in the HWE-based construction of an advanced C21–C34 fragment of the aplyronines. The strategic decision to incorporate the terminal *N*-methyl-*N*-vinyl formamide in subunit **5** with *cis*-geometry and to carry this potentially sensitive moiety through to the closing stages of the synthesis before isomerisation, contrasts with established approaches in which the required *trans*-alkenyl formamide is generally introduced by a testing, late-stage, condensation reaction with a highly functionalised aldehyde.^{2,7}

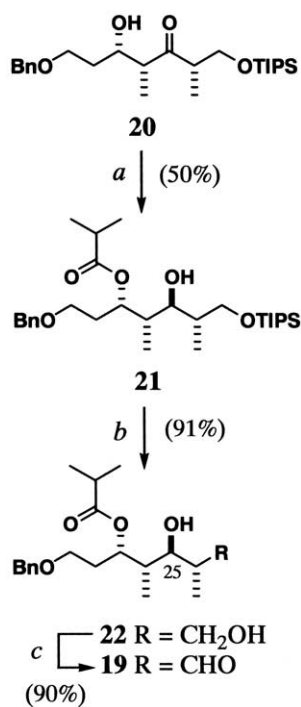
The synthesis of the C28–C34 phosphonate subunit **5**, containing the terminal *N*-methyl-*N*-vinyl formamide, is outlined in Scheme 2. Enolisation of lactate-derived ketone **6** using $(c\text{-Hex})_2\text{BCl}/\text{Me}_2\text{NEt}$ gave the (*E*)-boron enolate **7**.⁸ Addition of freshly prepared aldehyde **8** gave, on oxidative workup, *anti*-aldol adduct **9** in 94% yield with >95% diastereoselectivity. Notably, the high level of stereoinduction exerted by the enolate leads here to anti-Felkin attack on the α -chiral aldehyde. Treatment of **9** with TESOTf and 2,6-lutidine gave TES ether **10** (92%). Reduction by LiBH_4 then furnished 1,2-diol **11** which was oxidatively cleaved

with NaIO_4 to give aldehyde **12** (87%). Following our standard conditions,⁹ Wittig olefination of aldehyde **12** with ylide **13**, generated in situ by the addition of LiHMDS to phosphonium salt **14**, proceeded smoothly to give *cis*-alkene **15** in 72% yield. After PMB ether cleavage with DDQ to give **16**, the liberated C29 hydroxyl group was oxidised with catalytic TPAP and NMO ¹⁰ to give aldehyde **17** (80%). Addition of lithiated methyl dimethylphosphonate to aldehyde **17** gave the desired hydroxyphosphonate **18** in 72% yield. Subsequent oxidation, again using the TPAP/ NMO protocol, provided β -ketophosphonate **5** in nine steps and 17% yield from ketone **6**. Notably, the vinyl formamide was carried through this sequence of reactions without complication. In contrast to *trans*-*N*-methyl-*N*-vinyl formamides, which tend to exist as mixtures of rotamers in solution at ambient temperature, the corresponding *cis*-isomers appear as a single rotamer by ¹H NMR spectroscopy. As this facilitates analysis, we decided to retain the *cis*-geometry in **5** for as long as possible through the remainder of the synthesis.

In preparation for examining the key HWE coupling step, the C21–C27 aldehyde **19** was prepared from the previously described aldol adduct **20** (Scheme 3).^{5a} Subjection of β -hydroxyketone **20** to Evans–Tishchenko reduction conditions¹¹ gave the desired monoprotected 1,3-*anti* diol **21**. HF-pyridine mediated removal of the TIPS ether then gave diol **22** cleanly (91%). Subsequent selective oxidation of the primary alcohol using cata-



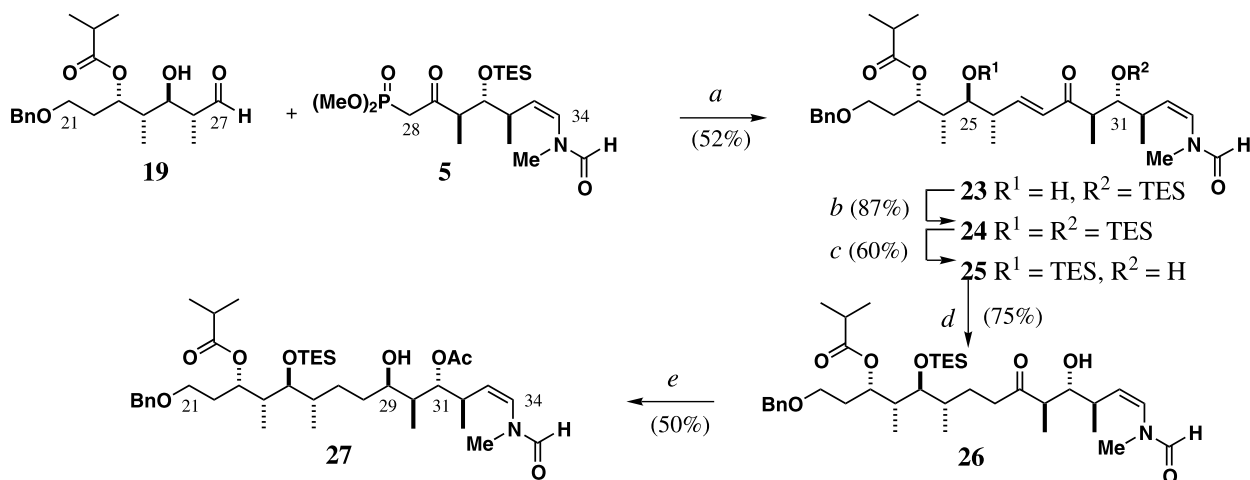
Scheme 2. (a) $(c\text{-Hex})_2\text{BCl}$, Me_2NEt , Et_2O , 0°C , 1 h; **8**, $-78 \rightarrow -20^\circ\text{C}$, 16 h; (b) TESOTf, 2,6-lutidine, CH_2Cl_2 , -78°C , 2 h; (c) LiBH_4 , THF, $-78^\circ\text{C} \rightarrow \text{rt}$, 24 h; (d) NaIO_4 , MeOH/pH 7 buffer, $0^\circ\text{C} \rightarrow \text{rt}$, 2 h; (e) LiHMDS , THF, $-78^\circ\text{C} \rightarrow \text{rt}$; **12**, -78°C , 2 h; (f) DDQ, $\text{CH}_2\text{Cl}_2/\text{pH}$ 7 buffer, rt , 0.5 h; (g) 10 mol% TPAP, NMO , 4 Å molecular sieves, CH_2Cl_2 , rt , 0.5 h; (h) $(\text{MeO})_2\text{POCH}_3$, $n\text{-BuLi}$, 4 Å mol. sieves, THF, -78°C , 4 h; (i) 10 mol% TPAP, NMO , 4 Å molecular sieves, CH_2Cl_2 , rt , 1 h.



Scheme 3. (a) *iso*-butyraldehyde, SmI₂ (20 mol%), THF, 0°C, 2.5 h; (b) HF·pyridine, THF, rt, 7 h; (c) cat. TEMPO, PhI(OAc)₂, rt, 2 h.

lytic TEMPO, with iodobenzene diacetate (BAIB) as a co-oxidant, as developed by Piancatelli and co-workers,¹² provided the desired aldehyde **19** (90%) without any oxidation at C25.

In order to form the C27–C28 bond of the aplyronines, the crucial HWE reaction between aldehyde **19** and β-ketophosphonate **5** was carried out using the LiCl/DBU conditions of Masamune and Roush,¹³ producing the (*E*)-enone **23** in 52% yield (Scheme 4). Subsequent attempts to protect the C25 alcohol as either a PMB or TBS ether proved unsuccessful, presumably due to the

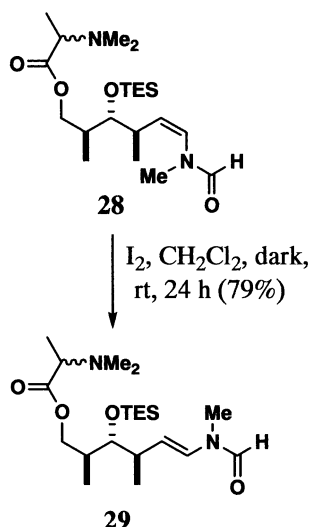


Scheme 4. (a) LiCl, DBU, THF/MeCN, rt, 3 h; (b) TESCl, imidazole, DMF, rt, 2 h; (c) AcOH/THF/H₂O, 40°C, 4 h; (d) [Ph₃P·CuH]₆, C₆H₆, rt, 1 h; (e) SmI₂, CH₃CHO, THF, –5°C, 2.5 h.

particularly hindered nature of this hydroxyl group. However, treatment of **23** with TESCl and imidazole in DMF gave the bis-TES ether **24** in high yield (87%). By using mild hydrolysis conditions (AcOH/THF/H₂O), selective deprotection of the C31 TES ether was achieved to give alcohol **25**. Stryker's reagent ([Ph₃P·CuH]₆) was then employed to reduce selectively the enone functionality in a 1,4-sense, providing ketone **26** (75%).¹⁴ Our synthesis of fragment **27**,¹⁵ corresponding to an advanced C21–C34 subunit of the aplyronines, was then concluded by performing an Evans–Tishchenko reduction on **26** using acetaldehyde. This served to set simultaneously the configuration of the C29 hydroxyl-bearing stereocentre and introduce the required C31 acetate.

Whilst retaining the *cis*-geometry of the terminal vinyl formamide through the above reaction sequence proved useful in simplifying the ¹H NMR spectra, ultimately it needs to be isomerised to the *trans*-geometry in the presence of the full aplyronine functionality. To date, this transformation has been demonstrated in a series of model substrates using iodine under light-free conditions.⁹ For example, the *cis*-vinylformamide **28** (prepared in 94% yield by acylation of **16** with (*R,S*)-*N,N*-dimethylalanine using DCC, DMAP, CH₂Cl₂) was isomerised cleanly to the corresponding *trans*-*N*-methyl-*N*-vinylformamide **29** in 79% yield (Scheme 5).

In conclusion, we have prepared the β-ketophosphonate **5** to be employed as a highly functionalised C28–C34 side chain coupling unit for the aplyronines. The synthesis features a boron-mediated *anti*-aldol reaction of the chiral ketone **6** to set up the stereochemistry and a novel Wittig olefination to introduce the *N*-methyl-*N*-vinylformamide moiety. An advanced C21–C34 fragment **27** for the aplyronines was then assembled by a testing HWE coupling between **5** and aldehyde **19** and elaboration of the resulting enone **23**. Studies towards completing a total synthesis of the aplyronines using this chemistry are currently under investigation.



Scheme 5.

Acknowledgements

We thank the EPSRC, the British Council (Commonwealth Scholarship to SBB), Merck Sharp & Dohme and Trinity College, Cambridge, for support.

References

1. Yamada, K.; Ojika, M.; Ishigaki, T.; Yoshida, Y.; Eki-moto, H.; Arakawa, M. *J. Am. Chem. Soc.* **1993**, *115*, 11020.
2. (a) Kigoshi, H.; Ojika, M.; Ishigaki, T.; Suenaga, K.; Mutou, T.; Sakakura, A.; Ogawa, T.; Yamada, K. *J. Am. Chem. Soc.* **1994**, *116*, 7443; (b) Kigoshi, H.; Suenaga, K.; Mutou, T.; Ishigaki, T.; Atsumi, T.; Ishiwata, H.; Sakakura, A.; Ogawa, T.; Ojika, M.; Yamada, K. *J. Org. Chem.* **1996**, *61*, 5326; (c) Suenaga, K.; Ishigaki, T.; Sakakura, A.; Kigoshi, H.; Yamada, K. *Tetrahedron Lett.* **1995**, *36*, 5053.
3. Saito, S.; Watabe, S.; Ozaki, H.; Kigoshi, H.; Yamada, K.; Fusetani, N.; Karaki, H. *J. Biochem.* **1996**, *120*, 552.
4. Gachet, Y.; Tournier, S.; Millar, J. B. A.; Hyams, J. S. *Nature* **2001**, *412*, 352.
5. (a) Paterson, I.; Cowden, C. J.; Woodrow, M. D. *Tetrahedron Lett.* **1998**, *39*, 6037; (b) Paterson, I.; Woodrow, M. D.; Cowden, C. J. *Tetrahedron Lett.* **1998**, *39*, 6041.
6. Marshall, J. A.; Johns, B. A. *J. Org. Chem.* **2000**, *65*, 1501.
7. For reviews on bioactive marine macrolide synthesis, see: (a) Norcross, R. D.; Paterson, I. *Chem. Rev.* **1995**, *95*, 2041; (b) Yeung, K.-S.; Paterson, I. *Angew. Chem., Int. Ed.* **2002**, *41*, in press.
8. (a) Paterson, I.; Wallace, D. J.; Velázquez, S. M. *Tetrahedron Lett.* **1994**, *35*, 9083; (b) Paterson, I.; Wallace, D. J. *Tetrahedron Lett.* **1994**, *35*, 9087; (c) Paterson, I.; Wallace, D. J.; Cowden, C. J. *Synthesis* **1998**, 639.
9. Paterson, I.; Cowden, C. J.; Watson, C. *Synlett* **1996**, 209.
10. Ley, S. V.; Norman, J.; Griffith, W. D.; Marsden, S. D. *Synthesis* **1994**, 639.
11. Evans, D. A.; Hoveyda, A. H. *J. Am. Chem. Soc.* **1990**, *112*, 6447.
12. De Mico, A.; Margarita, R.; Parlanti, L.; Vescovi, A.; Piancatelli, G. *J. Org. Chem.* **1997**, *62*, 6974.
13. Blanchette, M. A.; Choy, W.; Davis, J. T.; Essensfeld, A. P.; Masamune, S.; Roush, W. R.; Sakai, T. *Tetrahedron Lett.* **1984**, *25*, 2183.
14. Mahoney, W. S.; Brestensky, D. M.; Stryker, J. M. *J. Am. Chem. Soc.* **1988**, *110*, 291.
15. All new compounds gave spectroscopic data in agreement with the structures indicated. Compound **27** was isolated as a colourless oil: R_f 0.19 (40% EtOAc/40–60 pet. ether); IR (film) 3406 (s br, O-H), 2963 (s, C-H), 1734 (s, C=O), 1715 (s, C=O), 1684 (s, C=O), 1647 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ_{H} 8.15 (1H, s, NCHO), 7.33 (3H, m, ArH), 7.26 (2H, m, ArH), 5.98 (1H, d, $J=8.7$ Hz, C_1H), 5.25 (1H, dd, $J=10.7, 9.0$ Hz, C_2H), 5.16 (1H, m, C_{12}H), 4.80 (1H, dd, $J=10.2, 3.0$ Hz, C_4H), 4.47 (2H, ABq, $J=11.8$ Hz, ArCH_2O), 3.46 (2H, m, BnOCH_2), 3.40 (2H, m, C_6HOH , $\text{C}_{10}\text{HOTES}$), 3.02 (3H, s, NCH_3), 2.90 (1H, m, C_3H), 2.49 (2H, septet, $J=7.0$ Hz, $\text{C}_2\text{H}(\text{CH}_3)_2$, OH), 2.15 (3H, s, $\text{C}(\text{O})\text{CH}_3$), 1.94 (1H, m, $\text{C}_{13}\text{H}_A\text{H}_B$), 1.86 (1H, m, $\text{C}_{13}\text{H}_A\text{H}_B$), 1.77 (1H, m, C_{11}H), 1.68 (1H, m, C_5H), 1.55 (2H, obs, $\text{C}_8\text{H}_A\text{H}_B$, C_9H), 1.39 (3H, m, C_7H_2 , $\text{C}_8\text{H}_A\text{H}_B$), 1.13 (3H, d, $J=7.0$ Hz, $(\text{C}_3\text{H}_3)_A$), 1.13 (3H, d, $J=7.1$ Hz, $(\text{C}_3\text{H}_3)_B$), 1.06 (3H, d, $J=6.8$ Hz, C_3HCH_3), 0.93 (15H, ap t, $J=8.0$ Hz, C_9HCH_3 , $\text{C}_{11}\text{HCH}_3$, $\text{Si}(\text{CH}_2\text{CH}_3)_3$), 0.77 (3H, d, $J=6.9$ Hz, C_5HCH_3), 0.62 (6H, q, $J=8.0$ Hz, $\text{Si}(\text{CH}_2\text{CH}_3)_3$); ^{13}C NMR (100 MHz, CDCl_3) δ_{C} 176.6, 172.2, 162.4, 138.4, 128.8, 128.3, 127.7, 127.5, 124.7, 79.1, 78.6, 77.7, 73.0, 71.8, 67.5, 41.4, 38.7, 34.3, 32.1, 31.4, 28.9, 23.7, 23.0, 20.8, 19.1, 18.9, 17.4, 14.0, 10.9, 8.3, 7.1, 5.5; HRMS (ES^+) $[\text{M}+\text{Na}]^+$ found 728.4569, $\text{C}_{39}\text{H}_{67}\text{O}_8\text{NNaSi}$ requires 728.4534.